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## THE ACTIVITY OF ANTIOXIDANT ENZYMES AND THE EXPRESSION OF THEIR GENES IN THE RAT BRAIN AFTER INJECTING OF VINDEBURNOL IN THE MULTIPLE SCLEROSIS MODEL

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## АКТИВНОСТЬ АНТИОКСИДАНТНЫХ ФЕРМЕНТОВ И УРОВЕНЬ ЭКСПРЕССИИ ИХ ГЕНОВ В МОЗГЕ КРЫС ПРИ ВВЕДЕНИИ ВИНДЕБУРНОЛА В МОДЕЛИ РАССЕЯННОГО СКЛЕРОЗА

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### ABSTARCT

The article presents the results of a study on the effectiveness of vindeburnol, a structural analogue of the cerebral vasodilator vincamine, in the model of multiple sclerosis. The drug was injected subcutaneously in rats after modeling experimental allergic encephalomyelitis at a dose of 20 µg/kg, from the first manifestation of a neurological deficit over the next 10 days. In the somatosensory cortex of animals, the activity of superoxide dismutase, glutathione peroxidase and glutathione reductase, as well as the expression level of the SOD1, GPX4, GPX6 and GSR genes using real-time polymerase chain reaction were studied. It was found that vindeburnol does not affect the expression of the SOD1 gene, as well as the activity of superoxide dismutase, while increasing the expression of the GPS4, GPS6 and GRS genes, which determines the increased activity of glutathione peroxidase and glutathione reductase in rats in the model of multiple sclerosis.

### АННОТАЦИЯ

В статье представлены результаты исследования эффективности применения виндебурнола, структурного аналога церебрального вазодилатора винкамина, в модели рассеянного склероза. Препарат вводили подкожно крысам после моделирования экспериментального аллергического энцефаломиелита в дозировке 20 мкг/кг со дня первых проявлений неврологического дефицита в течение последующих 10 суток. В соматосенсорной коре животных исследовали активность супероксиддисмутазы, глутатионпероксидазы и глутатионредуктазы, а также уровень экспрессии генов SOD1, GPX4, GPX6 и GSR с использованием полимеразной цепной реакции в реальном времени. Установлено, что виндебурнол не оказывает влияния на экспрессию гена SOD1, как и на активность супероксиддисмутазы, при этом способствует повышению экспрессии генов GPS4, GPS6 и GRS, что определяет повышение активности глутатионпероксидазы и глутатионредуктазы у крыс в модели рассеянного склероза.

**Keywords:** multiple sclerosis, activity of antioxidative biocatalysts, gene expressions, vindeburnol.

**Ключевые слова:** рассеянный склероз, активность антиоксидантных ферментов, экспрессия генов, виндебурнол

Multiple sclerosis (MS) is a chronic inflammatory disease of the central nervous system that affects not only white, but also gray matter [1-2]. The histopathological feature of MS is the presence of foci

of demyelination, as a result of which severe neurological disorders are observed in patients. In the process of development of demyelination, the progenitor cells of microglia and oligodendrocytes

penetrate into the inflammation zone, initiating a spontaneous remyelination process. A distinctive feature of the neuroinflammatory process is a local increase in the intensity of free radical processes, which leads to the release of certain cytokines and chemokines by glial cells. This creates an environment for enhancing cellular interactions, which leads to the ability of oligodendrocytes to synthesize new myelin [3]. In the acute phases of MS, the formation of an excessive amount of reactive oxygen species (ROS) is observed, which suppress the activity of antioxidant enzymes, which, as a result, leads to oxidative stress. The intensification of free radical processes also contributes to damage to biological macromolecules, such as polyunsaturated fatty acids in membrane lipids, as well as proteins and nucleic acids. This is confirmed by the results of a study of free radical processes in the blood and cerebrospinal fluid of patients with multiple sclerosis during the active phases of the disease. This indicates that an increase in ROS levels could lead to depletion of cellular antioxidants and the development of damage [4-7].

It was previously found that in the model of multiple sclerosis, vindeburnol, a structural analogue of the cerebral vasodilator vincamine, reduces the activity of oligodendrocytes and the volume of demyelinated regions in mice, and also increases the expression level of a number of genes involved in the survival of Locus coeruleus cells [8].

This study presents the results of a study of the effect of vindeburnol on gene expression of antioxidant enzymes, as well as on the activity of these antioxidants in rat brain in a model of multiple sclerosis.

Research methods. Animal experiments were carried out in compliance with European Convention for the Protection of Vertebrate Animals Used for Experimental and other Scientific Purposes (Strasbourg, 18 March, 1986).

The experiment was conducted on outbred white female rats weighing 200-250 g, which were divided into groups: 1 – control group (n = 8); 2 – experimental allergic encephalomyelitis (EAE) was simulated in animals (n = 8); 3 – after simulation of EAE, vindeburnol was administered subcutaneously at a dose of 20 µg / kg from the first manifestations of neurological deficiency over the next 10 days (to the peak of the clinical manifestations of the disease). The calculation of the dose of vindeburnol was carried out by calculating the LD50 for a given animal species with a confidence interval of 95%. The spinal cord of outbred white rats without neurological symptoms and modified Freund's complete adjuvant served as material for immunizing animals to reproduce EAE. The reaction of animals to the administration of the drug was evaluated by the level of manifestation of neurological deficit and pathomorphological changes in the brain of rats of experimental groups at the peak of the acute phase of the disease; in this formulation of

an experiment on modeling EAE, signs of cerebral MS are manifested [9].

On the 30th day after the start of the experiment, the rats were decapitated; somatosensory cortex was isolated on ice, where free radical oxidation indices and gene expression levels (SOD1, GPX4, GPX6 and GSR) were studied.

Activity of glutathione peroxidase (GPx) in supernatant of hippocampus tissue was determined by speed of oxidation of reduced glutathione (GSH) in presence of tert-Butyl hydroperoxide according to the method [10]. Activity of glutathione reductase (GSR) was measured by speed of oxidation of NADPH oxidase according to the method [11]. The activities of GPx and GSR were expressed in µmol/min/mg protein. The activity of superoxide dismutase (SOD) in supernatants was determined by the intensity of inhibition of nitro blue tetrazolium reduction during superoxide radical generation during xanthine oxidation with xanthine oxidase at a wavelength of 560 nm, the enzyme activity was expressed in ΔE560 / mg protein [12].

The relative level of transcripts of the studied genes was investigated using the real-time polymerase chain reaction (Real-time PCR) method after reverse transcription. Total RNA from cortical and hippocampal tissues was isolated using the Aurum Total RNA fatty and fibrous tissue kit (BioRad). The amount and purity of the isolated RNA was evaluated spectrophotometrically using a NanoPhotometer (Implen). OD260/280 of RNA samples ranged from 1.80 to 2.00. Total RNA (0.5 µg) was reverse transcribed into cDNA, in a mixture volume of 25 µl. The reverse transcription reaction was carried out using reagents of the company Evrogen (Russia), in accordance with the manufacturer's instructions at 40°C for 40 minutes, and then inactivated at 92°C for 10 minutes. Design of forward and reverse primers for Real-time PCR was carried out in the Primer3 program with subsequent verification in IDT. The primers were synthesized by Synthol (Russia). The primer sequences are shown in table 1.

Real-time PCR was performed on a Bio-Rad CFX-96 amplifier using a ready-made iQ BioRad reaction mixture in a volume of 25 µl, including 50 pmol of forward and reverse primers and 1 µl of cDNA. Conditions for PCR: primary denaturation was carried out at 95°C for 3 minutes. 35 amplification cycles were carried out: 20s at 95°C, 10s at 60°C, 20s at 72°C. To verify the specificity after the end of the amplification cycles, a melting curve was constructed from 65°C to 95°C in increments of 0.5°C. SYBR Green I was used as a dye. The amplification reaction for each studied gene was carried out separately in duplicates. The HPRT1 gene encoding the hypoxanthine phosphoribosyltransferase 1 enzyme was used as the reference gene.

Table 1

Primer sequences			
Gene	Forward Primer	Reverse Primer	melting point (°C)
SOD1	Gcg-gat-gaa-gag-agg-cat-gtt	Acg-gcc-aat-gat-gga-atg-ct	60
GPX4	Tga-gcc-gct-tat-tga-agc-ca	Cac-acg-caa-ccc-ctg-tac-tt	60
GPX6	Acg-tac-cct-gaa-ctg-aac-aca	Ccg-ttc-aca-tcc-ccc-ttc-tc	60
GSR	Gcc-ttc-acc-ccg-atg-tat-ca	Gcc-aac-cac-ctt-ctc-ctc-ttt	60
HPRT1	Tcc-tcc-tca-gac-cgc-ttt-tc	Atc-act-aat-cac-gac-gct-ggg	60

Relative expression was calculated by evaluating the Ct (Threshold cycle) values obtained during Real-time PCR. Next, the expression levels of the studied genes in the experimental and control samples were compared using the  $2^{-\Delta\Delta C_t}$  method [13].

Statistical processing of the research results was performed using the ANOVA method and the Statistica 10.0. A check for the normality of the distribution of experimental data was carried out using the Kolmogorov-Smirnov and Livin test. Data are presented as mean values and standard errors. The differences were considered statistically significant at  $p < 0.05$ .

Results. Antioxidant enzymes (SOD, GPx, catalase) catalyze reactions with reactive oxygen species, leading to the formation of inactive products. There is a group of antioxidant enzymes that use reduced glutathione as a cofactor, electron and proton donor, and material for neutralizing electrophilic compounds. SOD neutralizes  $H_2O_2$  and lipid peroxides, showing its activity in protecting, mainly, membranes. GPR reduces oxidized glutathione, thereby reactivating the system. An extensive group of

glutathione-S-transferases, along with peroxidase activity, has the ability to alkylate toxic electrophilic compounds, conjugating them with HB-extra-erythrocyte hemoglobin. At the same time, the effect of detoxification is achieved, and GSH is irreversibly consumed [14]. Enzymes from the glutathione group, and also SOD, are predominantly present in the brain.

When studying the activity of antioxidant enzymes in the somatosensory cortex in the EAE model, a significant decrease in the activity was found: SOD by 59% ( $p < 0.05$ ), GPx by 37% ( $p < 0.05$ ) and GSR by 51% ( $p < 0.05$ ), relative to the control group. Under the conditions of the introduction of vindeburnol, the activity of these enzymes increased relative to the group of animals that were modeled by EAE. Nevertheless, the antioxidant defense link in these conditions did not correspond to the physiological norm: in animals in the EAE model and the introduction of vindeburnol, the activity of GSR, GPx and GSR was lower than the control level, respectively, by 32% ( $p < 0.05$ ), by 25% ( $p < 0.05$ ) and 35% ( $p < 0.05$ ) (Table 2).

Table 2

**The effect of vindeburnol on the activity of superoxide dismutase ( $\Delta E560/\text{mg protein}$ ), glutathione peroxidase ( $\mu\text{mol/min/mg protein}$ ) and glutathione reductase ( $\mu\text{mol/min/mg protein}$ ) in rat somatosensory cortex in the model of multiple sclerosis**

Groups	SOD	GPx	GSR
control	54,77±3,04	29,25±1,64	23,53±1,21
EAE model	22,31±1,19*	18,35±0,96*	11,62±0,07*
EAE model + vindeburnol	37,42±2,53*	21,84±1,29*	15,37±0,83*

\* – significant differences in indicators relative to the values in the control group (at  $p < 0.05$ )

Based on the results of the study of gene expression of antioxidant enzymes presented in Figure 1, the following conclusions can be made.

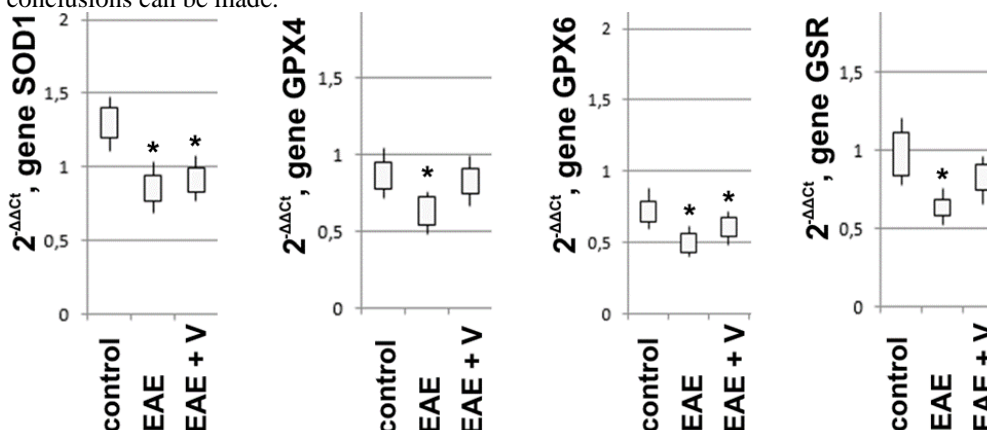


Fig. 1. The effect of vindeburnol on gene expression of antioxidant enzymes in rat somatosensory cortex in a model of multiple sclerosis. EAE - experimental allergic encephalomyelitis; V - vindeburnol

\* – significant differences in indicators relative to the values in the control group (at  $p < 0.05$ )

First of all, under the conditions of modeling multiple sclerosis in the somatosensory cortex of rats, a decrease in the expression of these genes is observed. The most significant decrease in the expression level was found for the SOD1 and GSR genes. Moreover, the introduction of vindeburnol contributes to an increase in the expression level of the GPS4, GRS genes and, to a lesser extent, the GPS6 gene. Vindeburnol has no effect on SOD1 gene expression.

Conclusion. It has now been proven that the neuropathology of multiple sclerosis is associated with the development of oxidative stress, excitotoxicity and inflammation. Studies on multiple sclerosis report an important role for SOD, which has neuroprotective properties in this pathology [15-17]. So, overexpression of Klotho in the mouse ALS SOD1 model leads to delayed onset and progression of the disease, increased survival. Klotho reduced the expression of neuroinflammatory markers and prevented the loss of neurons in the motor cortex and, especially, in the spinal cord [18]. The results obtained in this work showed that the use of vindeburnol in the model of multiple sclerosis has a greater effect on the glutathione system through its effect on the expression of GPS4, GRS genes and, to a lesser extent, the GPS6 gene; this drug does not affect the expression of SOD1. As a result, the activity of SOD is not compensated by the introduction of vindeburnol (in contrast to the enzymes GPx and GSR).

## References

1. Compston A, Coles A, Multiple sclerosis. *Lancet*. 2008;372:1502-1517. doi: 10.1016/S0140-6736(08)61620-7.
2. Hassanpour-Dehkordi A, Jivad N, Comparison of regular aerobics and yoga on the quality of life in patients with multiple sclerosis. *Med. J. Islam. Repub. Iran*. 2014;6:141.
3. Varas R, Ortiz FC, Neuroinflammation in Demyelinating Diseases: Oxidative Stress as a Modulator of Glial Cross-Talk. *Curr Pharm Des*. 2019;25(45): 4755-4762. doi: 10.2174/1381612825666191216125725.
4. Ortiz GG, Pacheco-Moisés FP, Bitzer-Quintero OK, et al., Immunology and oxidative stress in multiple sclerosis: Clinical and basic approach. *Clin. Dev. Immunol*. 2013;2013:708659. doi: 10.1155/2013/708659.
5. Van Horssen J, Witte ME, Schreibeit G, de Vries HE, Radical changes in multiple sclerosis pathogenesis. *Biochim. Biophys. Acta*. 2011;1812:141-150. doi: 10.1016/j.bbadis.2010.06.011.
6. Haider L, Fischer MT, Frischer JM, Bauer J, et al., Oxidative damage in multiple sclerosis lesions. *Brain*. 2011;134:1914-1924. doi: 10.1093/brain/awr128.
7. van Horssen J, Witte ME, Schreibeit G, de Vries HE, Radical changes in multiple sclerosis pathogenesis. *Biochim Biophys Acta*. 2011. 1812(2):141-50. doi: 10.1016/j.bbadis.2010.06.011.
8. Polak PE, Kalinin S, Braun D, et al. The vincamine derivative vindeburnol provides benefit in a mouse model of multiple sclerosis: effects on the Locus coeruleus. *Journal of neurochemistry*. 2012;121(2):206-216.
9. Карантыш Г.В., Гафиятуллина Г.Ш., Менджерский А.М. и др. Влияние виндебурнола на неврологический статус и морфологические изменения в головном мозге крыс в модели экспериментального аллергического энцефаломиелимита // Современные проблемы науки и образования. -2018. - № 1. <https://science-education.ru/ru/article/view?id=27398>. [Karantyshev GV. Gafiyatullina GSh. Mendzheritskiy AM. i dr. Vliyaniye vindeburnola na nevrologicheskiy status i morfologicheskiye izmeneniya v golovnom mozge krys v modeli eksperimentalnogo allergicheskogo entsefalomiyelita. *Sovremennyye problemy nauki i obrazovaniya*. 2018;1. <https://science-education.ru/ru/article/view?id=27398> (In Russ).].
10. Gunzler WA, Flohe L, Glutathione peroxidase *Handbook of methods for oxygen radical research*. Boca Ration: CRC Press, 1986;203-211.
11. Beutler E. Red cell metabolism: A Manual of Biochemical Methods. Grune and Stratton: New York, 1975;160.
12. Арутюнян А.В., Дубинина Е.Е., Зыбина Н.Н. Методы оценки свободнорадикального окисления и антиоксидантной системы организма: методические рекомендации. - СПб., 2005. – 208 с. [Arutyunyan AV. Dubinina EYe. Zybyina NN. Metody otsenki svobodnoradikalnogo okisleniya i antioksidantnoy sistemy organizma: metodicheskiye rekomendatsii. SPb. 2005;208. (In Russ).].
13. Livak KJ, Schmittgen TD, Analysis of relative gene expression data using real-time quantitative PCR and the 2(-delta delta C(T)) method. *Methods*. 2001;25:402-408.
14. Меньщикова Е.Б., Зенков Н.К., Ланкин В.З. и др. Окислительный стресс. Патологические состояния и заболевания. - Новосибирск: АРТА, 2008. – 284 с. [Menshchikova EB. Zenkov NK. Lankin VZ. i dr. Okislitelnyy stress. Patologicheskiye sostoyaniya i zabolevaniya. Novosibirsk: ARTA. 2008;284. (In Russ).].
15. Franklin RJ, French-Constant C, Edgar JM, Smith KJ, Neuroprotection and repair in multiple sclerosis. *Nat Rev Neurol*. 2012;8(11):624-634. doi: 10.1038/nrneurol.2012.200.
16. Hayashi Y, Homma K, Ichijo H. SOD1 in neurotoxicity and its controversial roles in SOD1 mutation-negative ALS. *Adv Biol Regul*. 2016;60:95-104. doi: 10.1016/j.jbior.2015.10.006.
17. Tokuda E, Marklund SL, Furukawa Y, Prion-like Properties of Misfolded Cu/Zn-superoxide Dismutase in Amyotrophic Lateral Sclerosis: Update and Perspectives. *Yakugaku Zasshi*. 2019;139(7):1015-1019. doi: 10.1248/yakushi.18-00165-5.
18. Zeldich E, Chen CD, Boden E, et al. Klotho Is Neuroprotective in the Superoxide Dismutase (SOD1G93A) Mouse Model of ALS. *J Mol Neurosci*. 2019;69(2):264-285. doi: 10.1007/s12031-019-01356-2.